



# **D-Xylose Microplate Assay Kit**

## **User Manual**

**Catalog # CAK1238**

(Version 2.2C)

Detection and Quantification of D-Xylose Content in Urine, Serum, Plasma, Tissue extracts, Cell lysate, Cell culture media, Other biological fluids Samples.

**For research use only. Not for diagnostic or therapeutic procedures.**

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## I. INTRODUCTION

In nature, D-xylose occurs mainly in the polysaccharide form as xylan, arabinoxylan, glucuronoarabinoxylan, xyloglucan and xylogalacturonan. Mixed linkage D-xylans are also found in certain seaweed species and a similar polysaccharide is thought to make up the backbone of psyllium gum. In humans, D-xylose is used in an absorption test to help diagnose problems that prevent the small intestine from absorbing nutrients, vitamins and minerals in food. D-Xylose is normally easily absorbed by the intestine. When problems with absorption occur, D-xylose is not absorbed and blood and urine levels are low. A D-xylose test can help to determine the cause of a child's failure to gain weight, especially when the child seems to be eating enough food. If, in a polysaccharide, the ratio of D-xylose to other sugars etc. is known, then the amount of the polysaccharide can be quantified from this knowledge plus the determined concentration of D-xylose in an acid hydrolysate. Xylans are a major portion of the polysaccharides that could potentially be hydrolysed to fermentable sugar for biofuel production.

D-Xylose Microplate Assay Kit provides a convenient tool for sensitive detection of D-Xylose in a variety of samples. D-xylose is oxidised by NAD<sup>+</sup> to D-xylonic acid in the presence of xylose dehydrogenase. D-Xylose is measured by the increase in absorbance at 450 nm.

## II.KIT COMPONENTS

Component	Volume	Storage
96-Well Microplate	1 plate	
Assay Buffer	30 ml x 4	4 °C
Reaction Buffer	10 ml x 1	4 °C
Coenzyme	Powder x 1	-20 °C
Enzyme	0.1 ml x 1	4 °C
Dye Reagent A	Powder x 1	4 °C
Dye Reagent B	1 ml x 1	4 °C
Standard	Powder x 1	4 °C
Plate Adhesive Strips	3 Strips	
Technical Manual	1 Manual	

**Note:**

**Coenzyme:** add 1 ml Reaction Buffer to dissolve before use.

**Enzyme:** add 1 ml Reaction Buffer to dissolve before use.

**Dye Reagent A:** add 9 ml distilled water to dissolve before use, mix, store at 4°C.

**Standard:** add 1 ml distilled water to dissolve before use; then add 0.05 ml into 0.95 ml distilled water, mix, the concentration will be 1 mmol/L.

### III. MATERIALS REQUIRED BUT NOT PROVIDED

1. Microplate reader to read absorbance at 450 nm
2. Distilled water
3. Pipettor, multi-channel pipettor
4. Pipette tips
5. Mortar
6. Centrifuge
7. Timer

### IV. SAMPLE PREPARATION

#### 1. For cell and bacteria samples

Collect cell or bacteria into centrifuge tube, discard the supernatant after centrifugation, add 1 ml Assay buffer for  $5 \times 10^6$  cell or bacteria, sonicate (with power 20%, sonicate 3s, interval 10s, repeat 30 times); centrifuged at 10000g 4°C for 15 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

#### 2. For tissue samples

Weigh out 0.1 g tissue, homogenize with 1 ml Assay buffer, centrifuged at 10000g 4°C for 15 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

#### 3. For serum or plasma samples

Detect directly.

## V. ASSAY PROCEDURE

Add following reagents into the microplate:

Reagent	Sample	Standard	Blank
Reaction Buffer	60 $\mu$ l	60 $\mu$ l	60 $\mu$ l
Sample	20 $\mu$ l	--	--
Standard	--	20 $\mu$ l	--
Assay Buffer	--	--	20 $\mu$ l
Coenzyme	10 $\mu$ l	10 $\mu$ l	10 $\mu$ l
Enzyme	10 $\mu$ l	10 $\mu$ l	10 $\mu$ l
Mix, cover the plate adhesive strip, put the plate into the convection oven, incubate at 37 °C for 10 minutes.			
Dye Reagent A	90 $\mu$ l	90 $\mu$ l	90 $\mu$ l
Dye Reagent B	10 $\mu$ l	10 $\mu$ l	10 $\mu$ l
Mix, measured at 450 nm immediately and record the absorbance.			

### Note:

- 1) Perform 2-fold serial dilutions of the top standards to make the standard curve.
- 2) The concentrations can vary over a wide range depending on the different samples. For unknown samples, we recommend doing a pilot experiment & testing several doses to ensure the readings are within the standard curve range.
- 3) Reagents must be added step by step, can not be mixed and added together.

## VI. CALCULATION

1. According to the weight of sample

$$\begin{aligned} \text{D-Xylose } (\mu\text{mol/g}) &= (C_{\text{Standard}} \times V_{\text{Standard}}) \times (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / (W \\ &\quad \times V_{\text{Sample}} / V_{\text{Assay}}) \\ &= (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / W \end{aligned}$$

2. According to the volume of sample

$$\begin{aligned} \text{D-Xylose } (\mu\text{mol/ml}) &= (C_{\text{Standard}} \times V_{\text{Standard}}) \times (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / \\ &\quad V_{\text{Sample}} \\ &= (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) \end{aligned}$$

$C_{\text{Standard}}$ : the concentration of standard, 1 mmol/L = 1  $\mu\text{mol/ml}$ ;

W: the weight of sample, g;

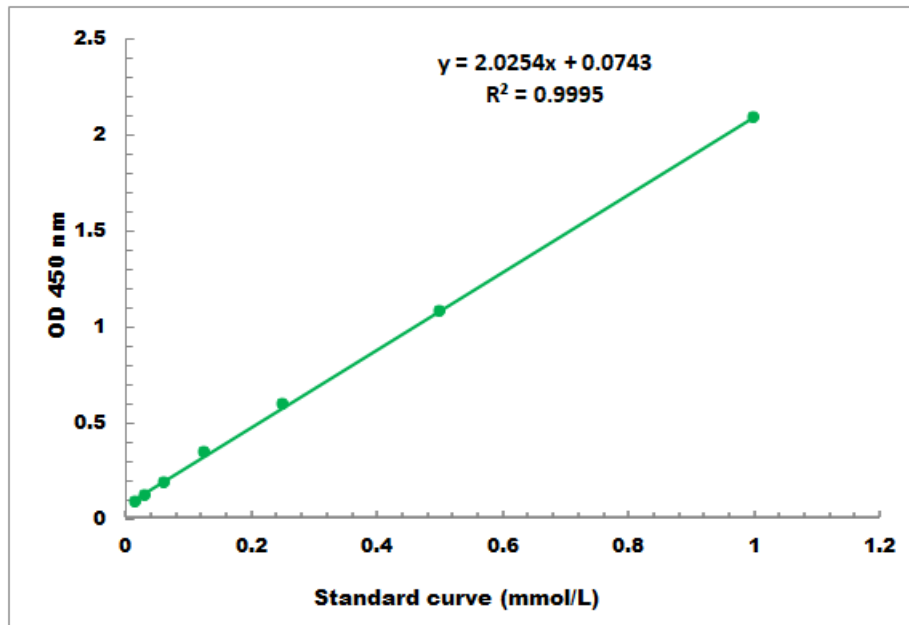
$V_{\text{Standard}}$ : the volume of standard, 0.02 ml;

$V_{\text{Sample}}$ : the volume of sample, 0.02 ml;

$V_{\text{Assay}}$ : the volume of Assay Buffer, 1 ml.

## VII. TYPICAL DATA

The standard curve is for demonstration only. A standard curve must be run with each assay.



Detection Range: 0.01 mmol/L - 1 mmol/L

## VIII. TECHNICAL SUPPORT

For troubleshooting, information or assistance, please go online to [www.cohesionbio.com](http://www.cohesionbio.com) or contact us at [techsupport@cohesionbio.com](mailto:techsupport@cohesionbio.com)

## IX. NOTES