

# **Trehalase Activity**

# **Fluorometric Microplate Assay Kit**

# **User Manual**

Catalog # CAK8007

(Version 1.1A)

Detection and Quantification of Trehalase (THL) Activity in Urine, Serum, Plasma, Tissue extracts, Cell lysate, Cell culture media and Other biological fluids Samples.

For research use only. Not for diagnostic or therapeutic procedures.



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## I. INTRODUCTION

Trehalase is a glycoside hydrolase enzyme that catalyzes the conversion of trehalose to glucose. It is found in most animals. It has been reported that more than 90% of total AT activity in S. cerevisiae is extracellular and cleaves extracellular trehalose into glucose in the periplasmic space.

Trehalase Microplate Assay Kit provides a simple and direct procedure for measuring trehalase activity in a variety of samples. This assay is initiated with the enzymatic hydrolysis of the trehalose by trehalase, resulting in the generation of an intermediate that reacts with the probe, which can be detected fluorometrically (Ex/Em 535/587).



### **II. KIT COMPONENTS**

Component	Volume	Storage
96-Well Black Microplate	1 plate	
Assay Buffer	30 ml x 4	4 °C
Reaction Buffer	20 ml x 1	4 °C
Substrate	Powder x 1	4 °C
Enzyme	Powder x 1	-20 °C
Probe	Powder x 1	-20 °C, keep in dark
Probe Diluent	1 ml x 1	4 °C
Standard (100 μmol/L)	1 ml x 1	4 °C
Positive Control	Powder x 1	-20 °C
Plate Adhesive Strips	3 Strips	
Technical Manual	1 Manual	

Note:

Substrate: add 1 ml Reaction Buffer to dissolve before use, mix. Store at 4 °C. Use within one month.

**Enzyme**: add 1 ml Reaction Buffer to dissolve before use. Aliquot & store at -20 °C. Use within one month.

Probe: Warm Probe Diluent to RT prior to use to melt frozen Probe Diluent; then add 1 ml Probe Diluent to dissolve. Store at -20 °C, protect from light and moisture. Use within one month.

**Positive Control**: add 1 ml Assay Buffer to dissolve before use. Store at -80 °C. Use within one month.



## III. MATERIALS REQUIRED BUT NOT PROVIDED

- 1. Fluorescence microplate reader to read fluorescence at Ex/Em = 535/587
- 2. Distilled water
- 3. Pipettor, multi-channel pipettor
- 4. Pipette tips
- 5. Mortar
- 6. Centrifuge
- 7. Timer
- 8. Ice

### IV. SAMPLE PREPARATION

1. For cell and bacteria samples

Collect cell or bacteria into centrifuge tube, discard the supernatant after centrifugation, add 1 ml Assay buffer for  $5 \times 10^6$  cell or bacteria, sonicate (with power 20%, sonicate 3s, interval 10s, repeat 30 times); centrifuged at 8000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

### 2. For tissue samples

Weigh out 0.1 g tissue, homogenize with 1 ml Assay buffer on ice, centrifuged at 8000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

For liquid samples
Detect directly.



# V. ASSAY PROCEDURE

Warm the solution to room temperature before use.

Add following reagents into the microplate:

Reagent	Sample	Control	Standard	Blank	Positive		
					Control		
Reaction Buffer	160 µl	160 µl	170 µl	170 µl	160 µl		
Sample	10 µl						
Distilled water		10 µl		10 µl			
Standard			10 µl				
Positive Control					10 µl		
Substrate	10 µl	10 µl			10 µl		
Probe	10 µl	10 µl	10 µl	10 µl	10 µl		
Enzyme	10 µl	10 µl	10 µl	10 µl	10 µl		
Mix, put it in the oven, 37 °C for 10 minutes, protected from light, record							

fluorescence measured at Ex/Em = 535/587 nm.

#### Note:

1) Perform 2-fold serial dilutions of the top standards to make the standard curve.

2) For unknown samples, we recommend doing a pilot experiment & testing several doses to ensure the readings are within the standard curve range. If the enzyme activity is lower, please add more sample into the reaction system; or increase the reaction time; if the enzyme activity is higher, please dilute the sample, or decrease the reaction time.

3) Reagents must be added step by step, can not be mixed and added together.



#### VI. CALCULATION

**Unit Definition:** One unit of trehalase activity is defined as the enzyme generates 1  $\mu$ mol H2O2 per min at 37° C.

1. According to the protein concentration of sample

THL (U/mg) = (C<sub>Standard</sub> × V<sub>Standard</sub>) × (OD<sub>Sample</sub> - OD<sub>Control</sub>) / (OD<sub>Standard</sub> - OD<sub>Blank</sub>) / (C<sub>Protein</sub> × V<sub>Sample</sub>) / T

= 0.01 × (OD<sub>Sample</sub> - OD<sub>Control</sub>) / (OD<sub>Standard</sub> - OD<sub>Blank</sub>) / C<sub>Protein</sub>

2. According to the weight of sample

THL (U/g) = (C<sub>Standard</sub> × V<sub>Standard</sub>) × (OD<sub>Sample</sub> - OD<sub>Control</sub>) / (OD<sub>Standard</sub> - OD<sub>Blank</sub>) / (V<sub>Sample</sub> × W/ V<sub>Assay</sub>) / T = 0.01 × (OD<sub>Sample</sub> - OD<sub>Control</sub>) / (OD<sub>Standard</sub> - OD<sub>Blank</sub>) / W

3. According to the quantity of cells or bacteria

 $THL (U/10^{4}) = (C_{Standard} \times V_{Standard}) \times (OD_{Sample} - OD_{Control}) / (OD_{Standard} - OD_{Blank}) / (V_{Sample} \times N / V_{Assay}) / T$  $= 0.01 \times (OD_{Sample} - OD_{Control}) / (OD_{Standard} - OD_{Blank}) / N$ 

4. According to the volume of sample

THL (U/mg) = (C<sub>Standard</sub> × V<sub>Standard</sub>) × (OD<sub>Sample</sub> - OD<sub>Control</sub>) / (OD<sub>Standard</sub> - OD<sub>Blank</sub>) / V<sub>Sample</sub> / T

= 0.01 × (OD<sub>Sample</sub> - OD<sub>Control</sub>) / (OD<sub>Standard</sub> - OD<sub>Blank</sub>)

 $C_{\text{Standard}}$ : the concentration of standard, 100  $\mu$ mol/L = 0.1  $\mu$ mol/ml;

C<sub>Protein</sub>: the protein concentration, mg/ml;

W: the weight of sample, g;

N: the quantity of cell or bacteria,  $N \times 10^4$ ;

V<sub>Standard</sub>: the volume of the standard, 0.01 ml;

V<sub>Sample</sub>: the volume of sample, 0.01 ml;

V<sub>Assay</sub>: the volume of Assay buffer, 1 ml;

T: the reaction time, 10 minutes.



### VII. TYPICAL DATA

The standard curve is for demonstration only. A standard curve must be run with each assay.



Detection Range: 1 µmol/L - 100 µmol/L



Positive Control reaction in 96-well plate assay with decreasing the concentration

#### VIII. TECHNICAL SUPPORT

For troubleshooting, information or assistance, please go online to www.cohesionbio.com or contact us at techsupport@cohesionbio.com

#### IX. NOTES