



**Beta-Lactamase Inhibitor Screening
Microplate Assay Kit
User Manual**

Catalog # CAK3001

(Version 1.1A)

Screening/studying/characterizing beta-Lactamase Inhibitors for efficient treatment of antibiotic resistant bacterial infections.

For research use only. Not for diagnostic or therapeutic procedures.

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I. INTRODUCTION

Beta-Lactamases (EC 3.5.2.6, β Ls), are a large family of hydrolases comprising more than 850 identified members expressed in Gram-positive and Gram-negative bacteria. β Ls can be classified according to their substrate or inhibitor specificity. These enzymes are capable of hydrolyzing four atom rings known as β -lactams. Antibiotics containing β -lactam rings (i.e. penicillin, cephalosporin, monobactam, carbapenem) are highly susceptible to be hydrolyzed via enzymatic activity, which deactivates their antibiotic potency. β Ls have become a significant clinical threat due to the alarming number of cases of bacterial strains showing β -lactam antibiotic resistance. β -Lactamase inhibition has become an urgent target in the treatment of bacterial infections displaying β -Lactam resistance. Several β -Lactam derivatives have been reported as β -Lactamase inhibitors; however, only clavulanic acid, sulbactam and tazobactam have reached clinical importance.

Beta-Lactamase Inhibitor Screening Microplate Assay Kit provides a rapid, simple, sensitive, and reliable test suitable for high-throughput screening of β -Lactamase inhibitors. This kit is based on the hydrolysis of Nitrocefin, a chromogenic cephalosporin, in the presence of Tazobactam—a potent inhibitor of β -Lactamase, the rate of hydrolysis of the substrate is decreased.

II. KIT COMPONENTS

Component	Volume	Storage
96-Well Microplate	1 plate	
Reaction Buffer	20 ml x 1	4 °C
Substrate	Powder x 1	-20 °C, keep in dark
Diluent	0.2 ml x 1	4 °C
Enzyme	Powder x 1	-20 °C, keep in dark
Inhibitor Control (Tazobactam)	Powder x 1	-20 °C
Technical Manual	1 Manual	

Note:

Substrate: Warm Diluent to RT prior to use, centrifuge the Substrate tube briefly, add 0.2 ml Diluent to dissolve before use and vortex, then add 0.8 ml Reaction Buffer mix. Store at -20 °C. Use within one month.

Inhibitor Control: Centrifuge the tube briefly, add 1 ml Reaction Buffer to dissolve before use and vortex, the concentration will be 5 mmol/L. (Dilute Inhibitor Control 5-fold by adding 4 µl of Inhibitor Control into 16 µl of Diluent). Store at -20 °C. Use within one month.

Enzyme: add 1 ml Reaction Buffer to dissolve before use, then add 20 µl into 980 µl Reaction Buffer mix. Aliquot & store at -80 °C. Use within one month.

III. MATERIALS REQUIRED BUT NOT PROVIDED

1. Microplate reader to read absorbance at 490 nm
2. Distilled water
3. Pipettor, multi-channel pipettor
4. Pipette tips
5. Centrifuge
6. Timer

IV. SAMPLE PREPARATION

Dissolve test inhibitors into proper solvent.

V. ASSAY PROCEDURE

Warm all reagents to room temperature before use.

Add following reagents into the microplate:

Reagent	Sample	Control	Inhibitor Control	Blank
Reaction Buffer	60 μ l	60 μ l	60 μ l	60 μ l
Sample	20 μ l	--	--	--
Distilled water	--	20 μ l	--	30 μ l
Inhibitor Control	--	--	20 μ l	--
Enzyme	10 μ l	10 μ l	10 μ l	--
Substrate	10 μ l	10 μ l	10 μ l	10 μ l
Mix, measure the absorbance in kinetic mode for 5-30 min. at 490 nm. Choose two time points (T1 & T2) in the linear range of the plot and obtain the corresponding values for the absorbance (Abs1 and Abs2).				

Note:

- 1) Perform 5-fold serial dilutions of the samples as much as needed.
- 2) The Control must be set up each time the assay is run.
- 3) Reagents must be added step by step, can not be mixed and added together.

VI. CALCULATION

$$\text{Slope}_{\text{Control}} = (\text{Abs2}_{\text{Control}} - \text{Abs1}_{\text{Control}}) / (T2 - T1)$$

$$\text{Slope}_{\text{Sample}} = (\text{Abs2}_{\text{Sample}} - \text{Abs1}_{\text{Sample}}) / (T2 - T1)$$

$$\text{Relative Inhibition (\%)} = (\text{Slope}_{\text{Control}} - \text{Slope}_{\text{Sample}}) / \text{Slope}_{\text{Control}} \times 100\%$$

$\text{Slope}_{\text{Sample}}$: the slope of the Sample Inhibitor

$\text{Slope}_{\text{Control}}$: the slope of the Enzyme Control

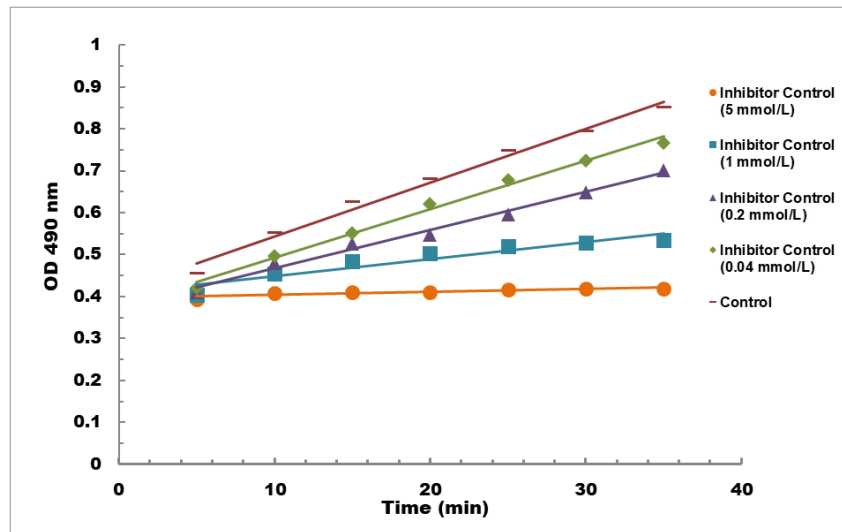
$\text{Abs1}_{\text{Sample}}$: the Sample Inhibitor absorbance at time point T1

$\text{Abs2}_{\text{Sample}}$: the Sample Inhibitor absorbance at time point T2

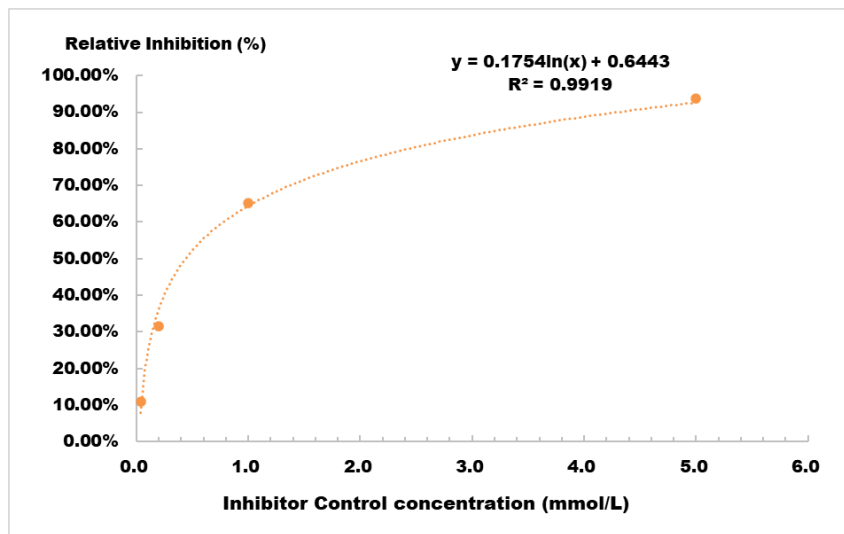
$\text{Abs1}_{\text{Control}}$: the Enzyme Control absorbance at time point T1

$\text{Abs2}_{\text{Control}}$: the Enzyme Control absorbance at time point T2

VII. TYPICAL DATA



Inhibition of β -Lactamase Enzymatic Activity with Tazobactam



Different concentrations inhibitor control (Tazobactam) relative inhibition

VIII. TECHNICAL SUPPORT

For troubleshooting, information or assistance, please go online to www.cohesionbio.com or contact us at techsupport@cohesionbio.com

IX. NOTES