

Malate Microplate Assay Kit User Manual

Catalog # CAK1187

(Version 1.1A)

Detection and Quantification of Malate content in Food, Juice,

Beverage, Other agricultural products Samples.

For research use only. Not for diagnostic or therapeutic procedures.



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I. INTRODUCTION

L-malate, or L-Malic acid, is a dicarboxylic acid that is made by all living organisms and plays an important role in the Calvin and Krebs Cycle. It is a source of CO2 for the Calvin cycle in plants and is also an intermediate that forms from fumarate in the Krebs Cycle. Malate is frequently used in food and beverage industries as an additive in products such as wine, beer, candies, etc.

Malate Microplate Assay Kit is designed to directly measure malate content in a variety of samples. It is based on malate dehydrogenase catalyzed oxidation of malate in which the formed NADH reduces a formazan reagent. The intensity of the product color, measured at 450 nm is proportional to the malate concentration in the sample.



II. KIT COMPONENTS

Component	Volume	Storage
96-Well Microplate	1 plate	
Reaction Buffer	10 ml x 1	4 °C
Enzyme	Powder x 1	-20 °C
Dye Reagent A	Powder x 1	4 °C
Dye Reagent B	1 ml x 1	4 °C
Standard	Powder x 1	4 °C
Plate Adhesive Strips	3 Strips	
Technical Manual	1 Manual	

Note:

- **Enzyme**: add 1 ml Reaction Buffer to dissolve before use. Aliquot & store at -80 °C. Use within one month.
- **Dye Reagent A**: add 1 ml distilled water to dissolve before use, mix. Store at 4 °C. Use within one month.
- Standard: add 1 ml distilled water to dissolve before use, then add 100 μ l into 400 μ l distilled water, the concentration will be 10 mmol/L. Store at -20 °C. Use within one month.



III. MATERIALS REQUIRED BUT NOT PROVIDED

- 1. Microplate reader to read absorbance at 450 nm
- 2. Distilled water
- 3. Pipettor, multi-channel pipettor
- 4. Pipette tips
- 5. Mortar
- 6. Centrifuge
- 7. Timer

IV. SAMPLE PREPARATION

1. For tissue samples

Weigh out 0.1 g tissue, homogenize with 1 ml distilled water, transfer it into microcentrifuge tube, centrifuged at 10000g for 10 minutes, take the supernatant into a new centrifuge tube for detection.

2. For liquid samples Detect directly.



V. ASSAY PROCEDURE

Add following reagents into the microplate:

Reagent	Sample	Standard	Blank	
Reaction Buffer	80 µl	80 µl	80 µl	
Sample	10 µl			
Standard		10 µl		
Distilled water			10 µl	
Enzyme	10 µl	10 µl	10 µl	
Dye Reagent A	90 μl	90 μl	90 μl	
Dye Reagent B	10 µl	10 µl	10 µl	
Mix, keep in dark for 30 minutes at room temperature, record absorbance				

measured at 450 nm.

Note:

1) Perform 2-fold serial dilutions of the top standards to make the standard curve.

2) The concentrations can vary over a wide range depending on the different samples.

For unknown samples, we recommend doing a pilot experiment & testing several

doses to ensure the readings are within the standard curve range.

3) Reagents must be added step by step, can not be mixed and added together.



VI. CALCULATION

1. According to the volume of sample

 $Malate (\mu mol/ml) = (C_{Standard} \times V_{Standard}) \times (OD_{Sample} - OD_{Blank}) / (OD_{Standard} - OD_{Blank}) /$

V_{Sample}

= (OD_{Sample} - OD_{Blank}) / (OD_{Standard} - OD_{Blank})

2. According to the weight of sample

Malate (µmol/g) = (C_{Standard} × V_{Standard}) × (OD_{Sample} - OD_{Blank}) / (OD_{Standard} - OD_{Blank}) / (V_{Sample} × W/ V_{Assay}) = (OD_{Sample} - OD_{Blank}) / (OD_{Standard} - OD_{Blank}) / W

 $C_{Standard}$: the standard concentration, 10 mmol/L = 10 μ mol/ml;

W: the weight of sample, g;

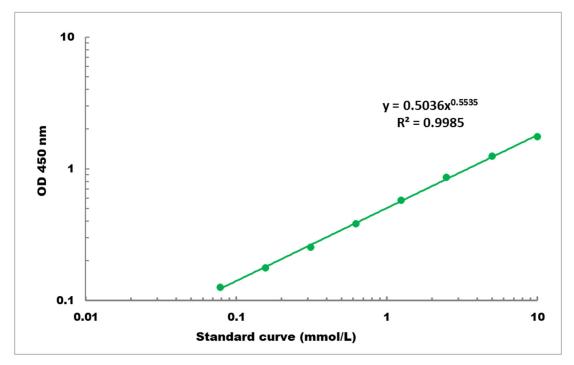
V_{Standard}: the volume of standard, 10 µl;

 V_{Sample} : the volume of sample, 10 µl.



VII. TYPICAL DATA

The standard curve is for demonstration only. A standard curve must be run with each assay.



Detection Range: 0.1 mmol/L - 10 mmol/L

VIII. TECHNICAL SUPPORT

For troubleshooting, information or assistance, please go online to www.cohesionbio.com or contact us at techsupport@cohesionbio.com

IX. NOTES