

Alkaline Phosphatase Microplate Assay Kit User Manual

Catalog # CAK1003

(Version 1.3E)

Detection and Quantification of Alkaline Phosphatase (ALP) Activity in Urine, Serum, Plasma, Tissue extracts, Cell lysate, Cell culture media and Other biological fluids Samples.

For research use only. Not for diagnostic or therapeutic procedures.



I. INTRODUCTION	2
II. KIT COMPONENTS	3
III. MATERIALS REQUIRED BUT NOT PROVIDED	4
IV. SAMPLE PREPARATION	4
V. ASSAY PROCEDURE	5
VI. CALCULATION	6
VII. TYPICAL DATA	7
VIII. TECHNICAL SUPPORT	7
IX NOTES	7



I. INTRODUCTION

Alkaline phosphatase (ALP) catalyzes the hydrolysis of phosphate esters in an alkaline environment, resulting in the formation of an organic radical and inorganic phosphate. In mammals, this enzyme is found mainly in the liver and bones. Marked increase in serum ALP levels, a disease known as hyperalkalinephosphatasemia, has been associated with malignant biliary obstruction, primary biliary cirrhosis, primary sclerosing cholangitis, hepatic lymphoma and sarcoidosis.

The assay is initiated with the enzymatic hydrolysis of the disodium phenyl phosphate by alkaline phosphatase. The enzyme catalysed reaction products can be measured at a colorimetric readout at 510 nm.



II. KIT COMPONENTS

Component	Volume	Storage
96-Well Microplate	1 plate	
Assay Buffer	30 ml x 4	4 °C
Reaction Buffer	4 ml x 1	4 °C, keep in dark
Substrate	Powder x 1	4 °C, keep in dark
Dye Reagent I	Powder x 1	4 °C, keep in dark
Dye Reagent II	Powder x 1	4 °C, keep in dark
Standard	Powder x 1	4 °C
Positive Control	0.1 μl x 1	4 °C
Plate Adhesive Strips	3 Strips	
Technical Manual	1 Manual	

Note:

Substrate: add 4 ml distilled water to dissolve before use.

Dye Reagent I: add 9 ml distilled water to dissolve before use.

Dye Reagent II: add 2 ml distilled water to dissolve before use.

Standard: add 1 ml distilled water to dissolve, then add 50 μ l standard into 950 μ l

distilled water, the concentration will be 5 mmol/L.

Positive Control: add 1 ml Assay Buffer to dissolve before use, mix.



III. MATERIALS REQUIRED BUT NOT PROVIDED

- 1. Microplate reader to read absorbance at 510 nm
- 2. Distilled water
- 3. Pipettor, multi-channel pipettor
- 4. Pipette tips
- 5. Mortar
- 6. Ice
- 7. Centrifuge
- 8. Timer

IV. SAMPLE PREPARATION

1. For cell and bacteria samples

Collect cell or bacteria into centrifuge tube, discard the supernatant after centrifugation, add 1 ml Assay buffer for 5×10⁶ cell or bacteria, sonicate (with power 20%, sonicate 3s, interval 10s, repeat 30 times); centrifuged at 8000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

2. For tissue samples

Weigh out 0.1 g tissue, homogenize with 1 ml Assay buffer on ice, centrifuged at 8000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

3. For serum or plasma samples

Detect directly, or dilute with Assay Buffer.



V. ASSAY PROCEDURE

Add following reagents in the microplate:

Reagent	Sample	Standard	Blank	Positive	
				Control	
Sample	10 μΙ				
Standard		10 μΙ			
Distilled water			10 μΙ		
Positive Control				10 μΙ	
Reaction Buffer	40 μΙ	40 μΙ	40 μΙ	40 μΙ	
Substrate	40 μΙ	40 μΙ	40 μΙ	40 μΙ	
Mix, put it in the oven, 37 °C for 15 minutes.					
Dye Reagent I	90 μΙ	90 μΙ	90 μΙ	90 μΙ	
Dye Reagent II	20 μΙ	20 μΙ	20 μΙ	20 μΙ	
Mix, wait for 10 minutes, record absorbance measured at 510 nm.					

Note:

- 1) Perform 2-fold serial dilutions of the top standards to make the standard curve.
- 2) For unknown samples, we recommend doing a pilot experiment & testing several doses to ensure the readings are within the standard curve range. If the enzyme activity is lower, please add more sample into the reaction system; or increase the reaction time; if the enzyme activity is higher, please dilute the sample, or decrease the reaction time.
- 3) Reagents must be added step by step, can not be mixed and added together.



VI. CALCULATION

Unit Definition: One unit of Alkaline Phosphatase activity is defined as the enzyme generates 1 μ mol phenol per minute.

1. According to the protein concentration of sample

ALP (U/mg) =
$$(C_{Standard} \times V_{Standard}) \times (OD_{Sample} - OD_{Blank}) / (OD_{Standard} - OD_{Blank}) / V_{Sample} / C_{Protein} / T$$

$$= 0.333 \times (OD_{Sample} - OD_{Blank}) / (OD_{Standard} - OD_{Blank}) / C_{Protein}$$

2. According to the weight of sample

ALP (U/g) =
$$(C_{Standard} \times V_{Standard}) \times (OD_{Sample} - OD_{Blank}) / (OD_{Standard} - OD_{Blank}) / (V_{Sample} \times W/V_{Assay}) / T$$

$$= 0.333 \times (OD_{Sample} - OD_{Blank}) / (OD_{Standard} - OD_{Blank}) / W$$

3. According to the volume of serum or plasma

ALP (U/mI) =
$$(C_{Standard} \times V_{Standard}) \times (OD_{Sample} - OD_{Blank}) / (OD_{Standard} - OD_{Blank}) / V_{Sample} / T$$

= 0.333 × $(OD_{Sample} - OD_{Blank}) / (OD_{Standard} - OD_{Blank})$

C_{Protein}: the protein concentration of sample, mg/ml;

W: the weight of sample, g;

 $C_{Standard}$: the concentration of standard, 5 mmol/L = 5 μ mol/ml;

V_{Standard}: the volume of standard, 0.01 ml;

V_{Sample}: the volume of sample, 0.01 ml;

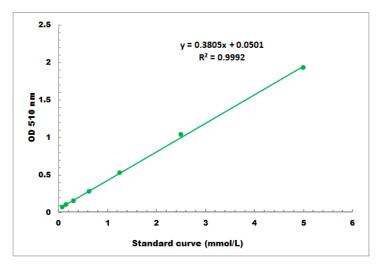
V_{Assay}: the volume of Assay buffer, 1 ml;

T: the reaction time, 15 minutes.

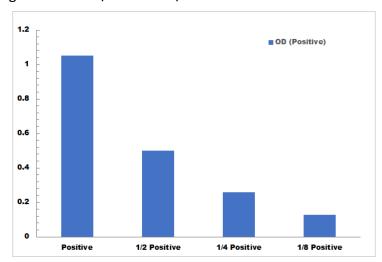


VII. TYPICAL DATA

The standard curve is for demonstration only. A standard curve must be run with each assay.



Detection Range: 0.04 mmol/L - 4 mmol/L



Positive Control reaction in 96-well plate assay with decreasing the concentration

VIII. TECHNICAL SUPPORT

For troubleshooting, information or assistance, please go online to www.cohesionbio.com or contact us at techsupport@cohesionbio.com

IX. NOTES